

Effects of flavonol-rich green tea (*Camellia sinensis* L. cv. Sofu) on blood glucose and insulin levels in diabetic mice

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Abstract

Findings from epidemiological studies and intervention trials seem to suggest that green tea consumption prevents type 2 diabetes. The anti-diabetic effects are most commonly attributed to the polyphenolic fraction of green tea. Green teas contain catechins; however, they also contain flavonols and their glycosides. Here, we compared between a common and a flavonol-rich tea cultivar, with regard to the anti-diabetic effects, in a type 2 diabetes mouse model using NSY (Nagoya-Shibata-Yasuda) mice. Water or a tea infusion, made from a common (control tea, TeaC) or a flavonol-rich (flavonol-rich tea, TeaF) cultivar, was given to the NSY mice as the solo drinking fluid from 4 until 13 weeks of age. At 12 weeks of age, fasting plasma insulin concentration was significantly lower in mice drinking TeaF than in mice drinking water. Relative to water, TeaC had no significant effect on plasma insulin levels. Thus, there were significant differences in efficacy with regard to maintenance of glucose homeostasis between a common and a flavonol-rich tea cultivars; therefore, tea flavonols are likely to have benefits with regard to preventing diabetes.

Introduction

Diabetes is serious public health issue, and its related conditions pose a growing and potentially enormous economic burden. Although effective surgical and pharmacological methods have been developed to treat symptoms related to diabetes, these treatments can be costly and are not without potential adverse effects [1-4]. The development of dietary agents for the prevention of diabetes could represent a cost-effective and safe approach to the public health problem. Epidemiological findings have indicated that green tea consumption may prevent type 2 diabetes, non-insulin dependent diabetes mellitus [5,6], and mounting evidence shows that green tea consumption improves glucose metabolism in human [7] and diabetic mice [8]. Green tea contains characteristic polyphenolic compounds, catechins which are thought to be primarily responsible for the physiological effects of green tea [9]. However, flavonols and their glycosides are also present in tea [10], and quercetin, one of the major tea flavonols, also has anti-diabetic effects in mouse models of diabetes [11-13]. We previously measured flavonol glycoside concentrations in tea infusions from various cultivars and identified cultivars with high flavonol contents [14]. Here, we compared the anti-diabetic effects between a common tea cultivar and a flavonol-rich cultivar to examine whether tea flavonols can ameliorate glucose intolerance in a mouse model of type 2 diabetes. Previously, NSY (Nagoya-Shibata-Yasuda) mice were generated by selective ICR mouse colony to establish a mouse model of type 2 diabetes [15]; these mice spontaneously develop renal lesions similar to diabetic nephropathy, but do not become severe obesity at any age of development [16]. These characteristics are similar to the pathophysiological features of human type 2 diabetes patients; therefore, NSY mice were used as a model of type 2 diabetes in the present study.

Materials and methods

NSY mice (4-week-old, male) were purchased from Hoshino Lab Animals Co. (Ibaraki, Japan). The mice were given a standard laboratory diet (CRF-1, Charles River Laboratories Inc. Wilmington, MA) in an environmentally controlled room (24°C, 60% relative humidity, a 12-h light/dark cycle). This study was approved by the Ethical Committee on Animal Experiments at the NARO Institute of Vegetable and Tea Science (No.H26-01), and all animal experiments were performed by following Law no. 105 and Notification no. 6 of the government of Japan.

'Sofu' green tea leaves (Flavonol-rich Tea: TeaF) were obtained from the NARO plantation in Makurazaki, Kagoshima, Japan, and control 'Yabukita' tea leaves (Control Tea: TeaC) were obtained from the NARO plantation in Kanaya, Shizuoka, Japan. The dried green tea leaves of each cultivars were steeped separately in cold water for one hour; aliquots of the filtrates were frozen until used in experiments. The stock solutions were diluted with water just before being given to mice and fresh drinking fluid was provided to all mice every two days. LC/MS was used to measure flavonol glycoside levels in the diluted tea infusions as described previously [14]. HPLC was used to measure catechin and caffeine levels as described previously [17].

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To investigate whether either tea infusion could affect glucose intolerance, TeaC and TeaF were given to NSY mice as the solo drinking fluid (TeaC and TeaF groups, respectively) for 8 weeks. Control NSY mice were given water (Water group) instead of tea infusions. Body weight and food consumption were measured once and twice a week, respectively. Glucose tolerance tests (GTTs) were performed by intraperitoneally injecting glucose (2 g/kg body weight) into mice that had fasted overnight and had been drinking the experimental fluids for 4 or 8 weeks; these animals were 8 or 12 weeks of age, respectively. Blood samples were obtained from the tail vein at 0, 30, 60, 120 and 240 min after the glucose injection. Blood glucose concentration was measured using a Glucose Pilot system (Iwai Chemical Co., Tokyo, Japan). For measurement of insulin, whole blood was collected from the tail vein at 0 and 30 min during i.p. GTT. Each blood sample was immediately centrifuged (3,000 xg, 5min, 4°C) to separate the plasma. Libs Mouse Insulin ELISA Kits (Shibayagi Co., Ltd., Gunma, Japan) were used according to the manufacturer's protocol to measure plasma insulin levels. At the end of the experiment, the mice were 13 weeks of age; they fasted overnight and were then anesthetized with isoflurane. A blood sample was collected into a heparinized tube from the infra-axillary artery of each anesthetized mouse. The pancreas, liver, and epididymal fat pads were quickly removed from each animal and weighed. Triglyceride Test Wako kits (Wako Pure Chemical Industries, Ltd., Osaka, Japan) were used to measure plasma triglyceride levels enzymatically.

All data obtained are expressed as the mean ± SEM. One-way factorial analysis of variance (ANOVA) and subsequent Scheffe's post-hoc test were used to compare means among groups. When data were not normally distributed, the Kruskal-Wallis test was applied. Differences with a *p* value less than 0.05 were considered significant. Notably, to determine the significance of the effects of tea infusions on the blood glucose and insulin levels (Figure 1), repeated measures ANOVA was performed, and then, if statistical significance was detected by this analysis, further statistical comparisons among the groups were conducted for each measurement time by one-way ANOVA and subsequent Scheffe's tests.

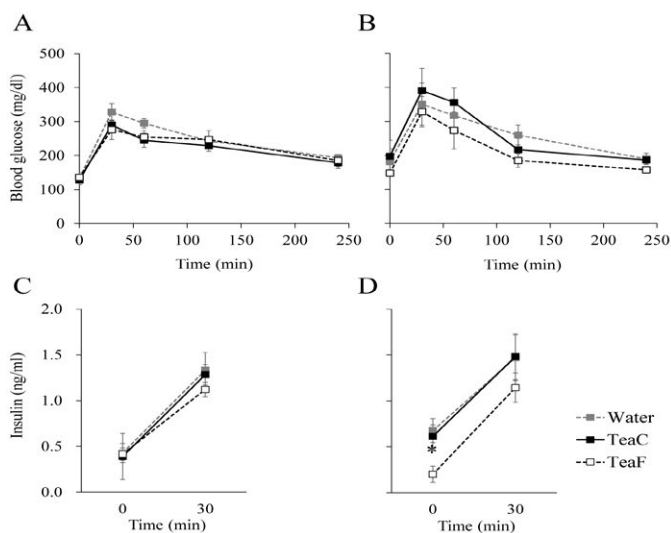


Figure 1. Effects of tea infusion on glucose tolerance and insulin response in NSY mice. Glucose tolerance tests (GTTs) were performed at 8 (A, C) or 12 (B, D) weeks of age. Blood glucose concentration (A, B) and plasma insulin (C, D) were measured during GTTs. All values are mean ± SEM (n = 4 or 5). **p*<0.05; significantly different from the Water group at each time-point, determined by one-way ANOVA, followed by Scheffe's test.

Results and discussion

Using protocols from our previous study [14], we determined the concentrations of two myricetin, five quercetin and three kaempferol glycosides in each type of tea infusion (Table 1). Among the measured quercetin glycosides, quercetin-3-*O*-glucosyl-rhamnosyl-glucoside was the most abundant in both tea infusions, TeaC and TeaF, as was the case in the previous study [14]. The level of quercetin-3-*O*-glucosyl-rhamnosyl-glucoside was notably higher in TeaF than in TeaC. Total aglycone content was also higher in TeaF than in TeaC for all of the aglycones measured. As shown in Table 2, while there were minor variations in the composition of catechins between TeaC and TeaF, the total amounts of catechins and caffeine in TeaF were similar to those in TeaC. Daily intakes of total flavonolglycones in TeaC and TeaF groups were approximately 700 and 1500 µg, respectively. For both tea infusion groups, daily intake of total catechins and caffeine were approximately 7.0 mg and 1.3 mg, respectively.

There were no significant differences among all groups with regard to body weight or food intake throughout the experiment (Table 3). Thus, relative to water, neither TeaC nor TeaF significantly affected body weight gain or food intake of NSY mice. The results of GTTs are shown in Figure 1. In the Water group, insulin response to glucose at 12 weeks of age (Figure 1B and 1D) was impaired relative to that at 8 weeks of age (Figure 1A and 1C). The result is consistent with previous findings that showed NSY mice spontaneously develop diabetes in an

Table 1. Flavonol glycoside levels in tea infusions.

Flavonol Glycosides (µg/ml)	TeaC	TeaF
Myricetin-3- <i>O</i> -glucoside	13.0	20.7
Myricetin-3- <i>O</i> -galactoside	29.8	44.6
Quercetin-3- <i>O</i> -glucoside (Isoquercitrin)	n.d	3.4
Quercetin-3- <i>O</i> -galactoside (Hyperoside)	6.1	15.1
Quercetin-3- <i>O</i> -rutinoside (Rutin)	1.4	20.6
Quercetin-3- <i>O</i> -glucosyl-rhamnosyl-glucoside	93.4	325.9
Quercetin-3- <i>O</i> -glucosyl-rhamnosyl-galactoside	80.0	90.4
Kaempferol-3- <i>O</i> -rutinoside	2.5	10.4
Kaempferol-3- <i>O</i> -glucosyl-rhamnosyl-glucoside	36.4	93.4
Kaempferol-3- <i>O</i> -glucosyl-rhamnosyl-galactoside	96.1	73.2
Total Aglycones (µg/ml)		
Myricetin	28.4	43.3
Quercetin	72.6	185.2
Kaempferol	51.3	68.1

Control Tea (TeaC) was made from a common tea cultivar, and Flavonol-rich Tea (TeaF) was made from a flavonol-rich tea cultivar. Total aglycone content was calculated from the concentrations of the corresponding glycosides. n.d.: not detected.

Table 2. Catechin and caffeine levels in tea infusions.

Catechins (mg/100 ml)	TeaC	TeaF
Catechin	1.9	3.2
Catechin gallate	n.d	0.2
Gallocatechin	2.9	4.4
Gallocatechin gallate	n.d	n.d
Epicatechin	16.6	25.4
Epicatechin gallate	5.5	5.4
Epigallocatechin	81.8	78.8
Epigallocatechin gallate	42.5	26.2
Total	151.2	143.6
Caffeine (mg/100 ml)	30.7	23.8

n.d: not detected

Table 3. Changes in body weight and food intake.

	Water	TeaC	TeaF
Body weight (g)			
4w	22.08 ± 1.08	21.46 ± 0.52	21.36 ± 0.38
8w	33.98 ± 1.21	33.76 ± 0.40	32.70 ± 0.63
12w	36.48 ± 1.47	35.70 ± 0.62	35.04 ± 0.39
Daily food intake (g/day/head)	4.31 ± 0.10	4.62 ± 0.18	4.52 ± 0.13

All values are mean ± SEM (n=4 or 5). There were no significant between-group differences in the body weight or food intake throughout the experiment.

Table 4. Effects of tea infusion on body weight, organ weight, and plasma triglyceride level in NSY mice.

	Water	TeaC	TeaF
Body weight (g)	32.83 ± 1.17	32.98 ± 0.53	32.34 ± 0.36
Pancreas (mg)	169.1 ± 7.0	168.5 ± 5.4	172.4 ± 7.4
Liver (g)	1.250 ± 0.029	1.298 ± 0.043	1.254 ± 0.034
Epididymal fat pad (mg)	1006.2 ± 135.3	726.6 ± 54.5	647.6 ± 48.8*
Plasma triglyceride (mg/dl)	135.0 ± 15.6	149.6 ± 9.3	119.7 ± 16.0

At 13 weeks of age, animals were denied food overnight and then subject to these assays in a fasting state. All values are mean ± SEM (n=4 or 5). * $p < 0.05$; significantly different from the Water group, determined by one-way ANOVA, followed by Scheffé's test.

age-dependent manner [16,18]. At 12 weeks of age, although glycemic excursions during the GTT were not significantly different among groups (Figure 1B), fasting plasma insulin concentration of the TeaF group was significantly lower than that of the Water group (Figure 1D). Fasting plasma insulin concentration of the TeaC group did not differ significantly from that of the Water group (Figure 1D). These result indicated that consumption of TeaF, but not TeaC, could improve the impaired insulin response in the NSY mice.

At the end of the experimental period, with the mice at 13 weeks of age and in a fasting state, there were no significant differences among the groups with regard to body weight, pancreatic weight, or liver weight (Table 4). However, epididymal fat pad weights were significantly less in the TeaF group than in the Water group. Mean fat pad weights in the TeaC group were also less than those in the Water group, however this difference was not significant ($p < 0.1$). The previous studies found mild obesity in the NSY mice [16,18], and mean fat pad mass of the Water group was greater than that of age-matched outbred ICR mice (484.1 ± 60.6 mg, n=4, unpublished data), from which the NSY mice were selectively bred for glucose intolerance. Therefore, these tea infusions, especially TeaF, are thought to suppress the fat accumulation in NSY mice. Numerous studies demonstrate that green tea and its extracts have anti-obesity effects in mouse models of diabetes [8,19]. For each of the three treatment groups of the NSY mice, fasting plasma triglyceride levels were distinctly higher than those of the age-matched ICR mice (85.5 ± 9.0 mg/dl, n=4, unpublished data) (Table 4). However, for NSY mice, there were no significant differences in the triglyceride levels among the three treatment groups (Water, TeaC, and TeaF); suggesting that TeaC and TeaF did not affect plasma triglyceride levels in NSY mice.

In the present study, we found that there were significant differences between a common and a flavonol-rich tea cultivars with regard to glucose homeostasis in NSY mice, and that tea flavonols are likely to have benefits for preventing diabetes. Further studies on tea flavonols and diabetes are warranted; such as those designed to obtain direct evidence of higher plasma levels of flavonols and their derivatives following consumption of flavonol-rich teas and examining the mechanisms in which flavonols improve insulin sensitivity in type 2 diabetic mouse models.

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